# Hairy Roots: From High-Value Metabolite Production to Phytoremediation

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### **ABSTRACT**

Environmental pollution is a global concern that is threatening the well-being of all life forms including humans. The cost of cleaning up contaminated sites is high and phytoremediation, the use of plants for removal of environmental pollutants, offers an attractive option due to its low cost and safety of implementation. The hairy roots technology has potential to become an excellent platform for studying numerous aspects encompassing phytoremediation. This is because hairy roots can be grown in large mass in culture media in a controlled environment and can therefore be subjected various physiological assays. Also. transformed roots are amenable to genetic manipulation and may facilitate the characterization of genes that influence the phytoremediation capacity of plants. This idea is well supported by the recent success in the development of transgenic plants for use in phytoremediation. Thus, hairy roots offer a good opportunity for the initial assessment of transgene efficacy in phytoremediation. Also, in the near future, hairy roots might be developed into initial screens for plants with enhanced capacity for phytoremediation. This review highlights the recent advances in the use of hairy roots to assess plants for their potential in removing important water and soil pollutants such as metals, explosives, radionuclides, insecticides, and antibiotics.

### Environmental pollution is a global concern

Environmental pollution is a global problem that affects both the developing and developed countries (Suresh and Ravishankar, 2004). To a large extent, both human and natural processes contribute to environmental pollution and contaminants are commonly classified as either organic or inorganic. Organic contaminants are a result of human activities including oil spills, military explosives, agriculture, fuel production, and wood treatment (Pilon-Smits, 2005). Common organic pollutants such as trichloroethylene (TCE), herbicides such as atrazine, explosives such as trinitrotoluene, petrochemicals such as benzene, toluene, polycyclic aromatic hydrocarbons, polychlorinated biphenyls

(PCBs), and the fuel additive methyl tert-butyl ether may contaminate soils and water (Xingmao and Burken, 2003; Pilon-Smits, 2005; Rentz et al., 2005; Suresh et al., 2005; González et al., 2006). In general, inorganic contaminants originate from either natural processes of soil weathering or human activities including agriculture and mining (Pilon-Smits, 2005). Subsequently, both natural and human activities may promote the release of heavy metals e.g. manganese, lead, copper, zinc, molybdenum, mercury, and nickel into soils and water posing a health threat to livestock and human populations (Nedelkoska and Doran, 2000a). For example, mercury is an important health concern to populations that rely heavily on the consumption of fish as a protein source (Hajeb et al., 2008), and to a large extent all global water bodies face the threat of mercury contamination (Harris et al., 2007).

### Plants are used to remove environmental contaminants

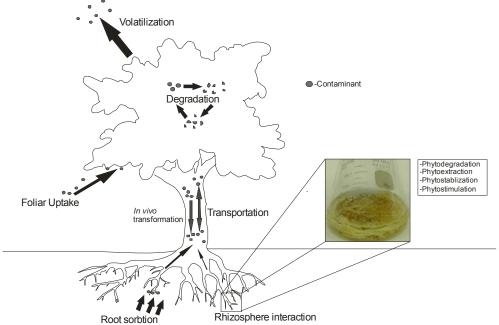
The health consequences due to environmental pollution are dire and the cost of cleaning up contaminated sites is high (Kuiper et al., 2004; Doty, 2008). Therefore, the use of plants to absorb, stabilize and degrade contaminants, collectively referred to as phytoremediation, is gaining acceptance as a more cost-effective alternative to other cleanup approaches. Phytoremediation is a technology that has been extensively reviewed (for recent reviews see Suresh and Ravishankar, 2004; Pilon-Smits 2005, and Doty, 2008). Our intention here is not to duplicate the efforts of the experts in the field, but instead we will concentrate this review on the potential of hairy roots as a powerful tool to study the phytoremediation capacity of plants.

The process of contaminant extraction by plants and the subsequent fates of the contaminant are described in Figure 1. Plant roots may act as a conduit for the absorption of a contaminant which is then translocated through the vascular system and concentrated in plant harvestable tissues in a process called phytoextraction (Doty, 2008). In addition, roots may provide a haven for microbial growth by secreting exudates that in turn act as a source of nutrition for the microbes and also serve as important cues for enhancing plant-microbe interactions (Bais et al., 2006). The resulting rhizospheric interactions may enhance the biodegradation of organic contaminants in a process referred to as

phytostimulation (Pilon-Smits, 2005 and references therein). Prior and after entering the plant via the root system, the contaminant may become target for degradation by either secreted or internal plant enzymes in a process called phytodegradation (Boominathan et al., 2004; Doty, 2008). The phytoremediation of some organic contaminants (e.g. TCE) is influenced by its concentration and the rate of transpiration, and TCE may be released from the plant through volatilization (Xingmao and Burken, 2003). Thus, in phytoremediation plants are used to facilitate optimum conditions for microbial break down of contaminants and to extract contaminants which may be metabolized or sequestered inside the plant (Boominathan et al., 2004; Tamaoki et al., 2005). Even though the rate of detoxification of organic contaminants in plant tissue is slow (Van Aken, 2008), the rising costs of physicochemical cleanup methods of contaminated sites makes phytoremediation a more attractive alternative (Doty, 2008 and references therein).

In order to mitigate the downward-migration of contaminants to the below-ground water reservoirs and lateral movement of contaminants via runoff and wind erosion, fast-transpiring

trees e.g. poplar (Populus sp.) are grown together with grasses resulting in phytostabilization of contaminants (Pilon-Smits, 2005). Therefore, in phytoremediation, plants provide dual benefits; they play the role of providing optimum conditions for root colonizing bacteria and also provide a simple and cost-effective way of extracting contaminants (Suresh and Ravishankar, 2004). Since roots are the primary contact between plant tissues and contaminants in the soil or water they provide a key point for assessment of the phytoremediation potential of a particular plant species. The underground portion of a plant system where roots are in contact with the micro biota is referred to as the rhizosphere (Walker et al., 2003) and the interaction among plant, microbes and mycorrhizal colonies is regulated to a large extent by root exudates (Walker et al., 2003; Bais et al., 2006). To that regard, root exudates are an essential component for pollutant degradation by microbes in the rhizosphere, and rhizosphere processes are thought to be essential for facilitating the uptake of contaminants by plants (Rentz et al., 2005). Therefore, the root environment and interactions among roots and microorganisms are key aspects to consider in phytoremediation (Barea et al., 2005).



**Figure 1.** Uptake and metabolism of environmental contaminants by plants: Contaminants can be absorbed by roots and foliage, transformed and degraded *in planta*, or volatilized into the atmosphere; rhizosphere interactions may also contribute to extraction and degradation of contaminants during phytoremediation. Hairy roots are a powerful tool to study various key processes that impact the overall phytoremediation capacity of plants, i.e. the rate of pollutant degradation, extraction, or stabilization. Hairy roots can also be used to study how root exudates may stimulate the degradation of particular contaminants.

## Hairy roots biotechnology for valuable metabolite production

Hairy roots are fine fibrous structures that are formed on plant tissues infected by *Agrobacterium rhizogenes*, a soil bacterium responsible for the root mat disease (Georgiev et al., 2007; Veena and Taylor, 2007). After infecting the cells, *A. rhizogenes* 

stably transfers several of its genes to the plant genome resulting in physiologic changes in the host cell leading to enhanced growth in hormone-free media (Srivastava and Srivastava, 2007). The observed changes in root physiology and morphology are associated with the transfer of a cluster of genes from the *A. rhizogenes* large Ri (root-inducing) plasmid into the plant genome. The symptoms

observed with *A. rhizogenes* infection may suggest that the transformed cells have been rendered more sensitive to auxin without altering the production of these plant hormones (McAfee et al., 1993; Srivastava and Srivastava, 2007).

Humankind has tapped into the plant natural products reservoir not only for nutritional needs, but also for medicinal and aesthetic purposes (Srivastava and Srivastava, 2007). However, to a high degree most valuable plant natural products are produced in small amounts from specialized metabolic pathways that fluctuate with respect to environmental conditions. The versatility of the hairy roots system has allowed the development of platforms for the production of high-value natural products, at times in scaled up bioreactors (Georgiev et al., 2007; Cuello and Yue, 2008; Villarreal et al., 2008; Weathers et al., 2008). In addition, the inherent characteristics of hairy roots including their fast growth, genetic stability, short doubling time, and ability to produce a broad range of metabolites similar to wild type make this system a powerful tool for metabolic engineering (Veena and Taylor, 2007). In combination with transgenic approaches, the capacity of hairy roots metabolism can be manipulated for the enhancement of de novo synthesis of high value phytochemicals (Guillon et al., 2006).

### Hairy roots technology offers important advantages for phytoremediation studies

Hairy roots offer several advantages for use in phytoremediation studies, these include: their ability to grow rapidly in microbe-free conditions, providing a greater surface area of contact between contaminant and tissue, and they are genetically and metabolically more stable in comparison to wild type (Gujarathi et al., 2005; Georgiev et al., 2007). Hairy roots are also amenable to genetic transformation, making gene transfer and characterization possible in a system that may pose minimum health or environmental concerns. Another advantage of using hairy roots for studying phytoremediation is their ability to produce large quantities of exudates which are composed of enzymes and some metal chelating compounds that may detoxify or sequester harmful organic and inorganic contaminants (Gujarathi et al., 2005; Bais et al., 2006; Doty, 2008). As shown in Table 1, hairy roots have been used to assess the potential of several plant species to remove contaminants from the environment. For example, the hairy root cultures of black nightshade (Solanum nigrum) may metabolize and remove PCBs from solutions spiked with PCB congeners (Macková et al., 1997a,b; Kučerova et al., 2000; Rezek et al., 2007). Also, by studying the rates of removal and the fate of contaminants such as the explosives hexahydro-1,3-5-trinitro-1,3-5-triazine (RDX) and octahydro-1,3,5,7tetranitro-1,3,5,7-tetrazocine (HMX), Badhra et al. (2001) discovered that periwinkle (*Catharanthus roseus*) hairy roots have an "intrinsic ability" to remove these molecules from the medium. RDX and HMX are the two most common pollutants found in military sites where explosives are commonly tested (Pilon-Smits, 2005).

Recently, hairy roots have been used to test plants for their ability to tolerate high levels of phenols (de Araujo et al., 2002). Phenols are commonly used in various agricultural applications or released from coal and petroleum refining activities, and they pose a threat to human health (de Araujo et al., 2002; Agostini et al., 2003; Coniglio et al., 2008). In hairy roots of carrot (Daucus carota) and other plant species the role of peroxidase enzymes might be the key factor in the removal of phenol and chlorophenols from the culture medium (Agostini et al., 2003; González et al., 2006; de Araujo et al., 2006; Singh et al., 2006; Coniglio et al., 2008). Also, the inherent activity of peroxidases in hairy roots of rapeseed (Brassica napus) was associated with the effective removal of 2,4-dichlorophenol and phenol from the medium for several cycles and the removal process was enhanced by exogenously-applied hydrogen peroxide (Agostini et al., 2003; Coniglio et al., 2008). It appears that other plants use additional mechanisms to remove phenol. For instance, cells of carrot, kangaroo apple (Solanum aviculare) and sweet potato (Ipomoea batatas) hairy roots are able to incorporate and conjugate phenolic compounds with polar cellular materials (possibly sugars and proteins) as well as with insoluble materials such as cell walls and membranes (de Araujo et al., 2006).

To a greater extent, the ability of plants to metabolize contaminants will depend on the biochemical characteristics of metabolizing enzymes and other protective mechanisms that may prolong tissue survival. Indeed, results from a comparative study of peroxidase enzymes from hairy roots of carrots, sweet potato and kangaroo apple demonstrated an inter-specific variation in the preference for phenol and chlorophenol among peroxidases (de Araujo et al., 2004). Also, peroxidase isozymes involved in phenol removal within a species may show variation in substrate preference and catalytic efficiency of phenol metabolism (Coniglio et al., 2008). It is noteworthy that, these studies are important in establishing an understanding of the enzymatic mechanisms of contaminant degradation for the selection of candidate enzymes that might be produced in large amounts and used as catalysts for contaminant break down (González et al., 2006).

An inspiring study by Eapen et al. (2003) demonstrated that hairy roots of the Indian mustard (*B. juncea*) and *Chenopodium amaranticolor* could remove uranium from solutions and could withstand

high concentrations of this radionuclide for days. It is encouraging to imagine that in the near future it may become possible to use plants to cleanup sites contaminated with radioactive waste and alleviate the devastating environmental problems that may arise through uranium contamination of soils and water (Gavrilkescu et al., 2008).

The uptake of metals and their distribution in plant tissues are both important aspects governing the capacity of plants to remove heavy metals from the soil. Hairy roots have demonstrated that they can be used as a means for screening a wide variety of plant species for their capacity to extract and sequester metals (Nedelkoska and Doran, 2000a). A comparative assessment of nickel tolerance between hairy roots and whole plants revealed that the translocation of nickel to above ground shoots may

not be required for nickel tolerance and hyperaccumulation in certain species of *Alyssum* (Nedelkoska and Doran, 2001). This suggests that nickel tolerance may be conferred by a reduced oxidative damage of hairy roots tissue due to enhanced catalase activity (Boominathan and Doran, 2002). Therefore, additional mechanisms to metal translocation and accumulation in shoots of hyperaccumulators may play a significant role in heavy metal tolerance. Indeed, using hairy roots, Boominathan and Doran (2003a) demonstrated that cadmium was extracted by alpine pennygrass (*Thlaspi caerulescens*) and accumulated in high levels in complexes with organic acids inside the cell walls.

**Table 1.** Phytoremediation of various environmental pollutants by hairy root cultures as tools to study the uptake and degradation of xenobiotics

Plant species	Model pollutant	Reference
Black nightshade (Solanum nigrum)	PCBs	Macková et al. (1997a; b)
Alpine pennygrass (Thlaspi caerulescens)	Cadmium	Nedelkoska and Doran (2000b)
Alyssum sp.	Nickel	Nedelkoska and Doran (2001)
Periwinkle (Catharanthus roseus)	RDX and HMX	Bhadra et al. (2001)
Carrot (Daucus carota)	Phenol and chloroderivatives	de Araujo et al. (2002)
Wild mustard (Alyssum bertolonii) and	Nickel, and cadmium	Boominathan and Doran (2002)
alpine pennygrass (T. caerulescens)		
Deadly nightshade (Atropa belladonna)	TCE	Banerjee et al. (2002)
Rapeseed (Brassica napus)	2,4-Dichlorophenol	Agostini et al. (2003)
Indian mustard ( <i>Brassica juncea</i> ) and Chenopodium amaranticolor	Uranium	Eapen et al. (2003)
Indian mustard (B. juncea) and chicory (Cichorium intybus)	DDT	Suresh et al. (2005)
Sunflower (Helianthus annuus)	Tetracycline and oxytetracycline	Gujarathi et al. (2005)
Tomato (Lycopersicon esculentum)	Phenols	Oller et al. (2005)
Carrot ( <i>D. carota</i> ), sweet potato ( <i>Ipomoea batatas</i> ), and kangaroo apple ( <i>Solanum aviculare</i> )	Guaiacol, catechol, phenol, 2- chlorophenol, and 2,6- dichlorophenol	de Araujo et al. (2004; 2006)
Indian mustard (B. juncea)	Phenol	Singh et al. (2006)
Tomato (L. esculentum)	Phenol	Wevar-Oller et al. (2005); González et al. (2006)
Rapeseed (B. napus)	Phenol	Coniglio et al. (2008)
Yellow tuft (Alyssum murale)	Nickel	Vinterhalter et al. (2008)

DDT= Dichloro-diphenyl-trichloroethane;, HMX=oxtahydro-1,3,5,7-tetranitro-1,3,5,7-tetrazocine; PCBs = polychlorinated biphenyls; RDX=hexahydro-1,3-5-trinitro-1,3-5-triazine; TCE=Trichloroethylene;

Also, in another study, Boominathan and Doran (2003b) revealed that an inherent high catalase activity may play an important role in cadmium hyperaccumulation in *T. caerulescens* hairy roots. Therefore, the establishment of hairy root cultures for a variety of plant species might be a good strategy in studies of growth and heavy metal tolerance in plants

(Nedelkoska and Doran, 2000b). Ultimately, the application of tissue culture technology may prove powerful in the regeneration of shoot cultures from hairy roots of selected species of plants with superior phytoremediation traits (Vinterhalter et al., 2008).

It is important to monitor and limit the release of pesticides and antibiotics into the environment, and of equal importance is the identification of methods for cleanup in the case of contamination. Hairy roots of sunflower (Helianthus annuus) are effective in extracting and metabolizing antibiotics including tetracycline and oxytetracycline through a process that is thought to involve reactive oxygen intermediates (Gujarathi and Linden, 2005). There is controversy regarding the continuous use of the insecticide DDT to combat mosquitoes that spread malaria in developing countries (Sadasivaiah et al., 2007) even though some studies suggest that DDT might have negative health effects on human health (Hatcher et al., 2008). Hairy roots of chicory (Cichorium intybus) and Indian mustard (Brassica juncea) have been used to study their potential in removing DDT from contaminated sites (Suresh et al... 2005). Interestingly, C. intybus and B. juncea might produce enzymes that degrade DDT (Suresh et al., 2005), thus offering a promising possibility for the characterization of these enzyme(s) and for similar studies to be done in other plant species.

The expression of heterologous proteins in hairy roots has successfully been done (Banerjee et al., 2002). Such an approach was used to express a mammalian cvtochrome P450 enzyme in deadly nightshade (Atropa belladonna) and the transgenic plants were able to metabolize the environmental pollutant TCE (Bernejee et al., 2002). Five years later, Doty et al. (2007) were successful in transforming poplar (Populus tremula x Populus alba) with this mammalian enzyme to generate plants with a superior capacity to remove various organic pollutants from hydroponic solutions and air. Of the several lines transformed with the mammalian enzyme, line 78 metabolized TCE a hundred-fold more than nontransgenic control trees (Doty et al., 2007). Also, others have used transgenic approaches that involved the over-expression of plant genes encoding contaminant metabolizing enzymes in hairy roots. For example. by over-expressing a tomato (Lycopersicon esculentum) tpx1 gene encoding a peroxidase in hairy roots, Wevar-Oller et al. (2005) generated roots with enhanced capacity of removing phenol from the medium. These studies demonstrated that transgenic approaches may be adopted to produce plants with novel and improved phytoremediation capacity (Van Aken, 2008). Therefore, in the near future the use of transgenic hairy root systems may become more

common in testing the efficacy of transgenes and the enzymes they encode for the removal of hazardous environmental pollutants.

All these studies demonstrate the power of using hairy roots in screening for candidate genes involved in the metabolism of environmental contaminants. Figure 2 illustrates a model of the mechanism(s) by which wild type or transgenic hairy root cells may metabolize environmental contaminants. It is noteworthy, however, that although the generation of transgenic plants with enhanced phytoremediation capacity might seem as a plausible solution, public skepticism and resistance to transgenic organisms might make this option less favorable for application in the near future. Alternatively, the selection of local plant species with enhanced phytoremediation capacity through hairy root screens may become more favorable and practical in the immediate future.

### **CONCLUSIONS AND FUTURE DIRECTIONS**

Hairy roots can be generated from many plant species by infecting them with A. rhizogenes. This technology has facilitated a more stable production of important medicinal and high-value products at times in scaled up bioreactors. The versatility of hairy roots makes this system more attractive for the assessment of various physiological aspects of plants. The problem of environmental pollution affects both local and global human populations and physicochemical technologies of environmental cleanup are costly. Therefore, the use of plants in phytoremediation is gaining more support. Plants have intrinsic abilities to extract and metabolize contaminants and their with soil microorganisms cooperation endophytes, microbes that live inside plants, may enhance the removal of contaminants from the environment. However, it is conceivable that not all species will possess superior capacities to extract and metabolize pollutants. These valuable plant traits can be screened for using hairy root cultures. Thus, the initial selection of superior plant species for use in phytoremediation can begin in the laboratory followed by the actual growing and testing plants in the greenhouse and the field. As hairy roots are amenable to genetic transformation, transgenic approaches may be used to study candidate genes that affect pollutant removal.

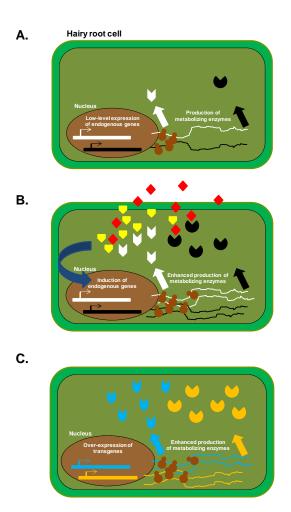


Figure 2. Metabolism of environmental contaminants by hairy root cells: (A) a cartoon depiction of a hairy root cell expressing contaminant metabolizing enzymes (white chevron and black pie) at basal levels; (B) environmental contaminants (red diamonds) may promote the production of reactive oxygen species (yellow pentagon), the enhanced production of ROS scavenging enzymes and antioxidants (white chevron), and/or contaminant metabolizing enzymes (black pie); (C) the expression of transgenes of animal or plant origin may also result in the enhanced production of contaminant metabolizing enzymes (blue chevron and orange pie) and phytoremediation capacity of plants.

Therefore, in the near future the hairy roots technology might be used more commonly in biotechnological efforts ranging from metabolite production to phytoremediation. Despite the large potential of hairy roots in phytoremediation studies, the ongoing challenge will be the actual translation of laboratory results to field applications. The lack of microbes in axenic hairy roots media may prevent our full appreciation of the benefits of the rhizospheric organisms that often enhance the uptake and breakdown of pollutants. Nevertheless, it is encouraging to witness the recent development of transgenic plants, poplar trees in particular, that promise to offer a tremendous impact on phytoremediation. In summary, hairy roots provide a

promising tool in the field of phytoremediation but the work of environmental remediation has just begun.

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### **REFERENCES CITED**

Agostini E, Coniglio MS, Milrad SR, Tigier HA, Giulietti AM (2003) Phytoremediation of 2,4-dichlorophenol by *Brassica napus* hairy root cultures. Biotechnol Appl Biochem 37: 139-144

Bais HP, Weir TL, Perry LG, Gilroy S, Vivanco JM (2006) The role of root exudates in rhizosphere interactions with plants and other organisms. Annu Rev Plant Biol 57: 233-266

Barea J, Pozo JM, Azcón R, Azcón-Aguilar C (2005) Microbial cooperation in the rhizosphere. J Expt Bot 56: 1761-1778

Bernejee S, Shang TQ, Wilson AM, Moore AL, Strand SE, Gordon MP, Doty SL (2002) Expression of functional mammalian P450 2E1 in hairy root cultures. Biotechnol Bioeng 77: 462-466

Bhadra R, Wayment DG, Williams RK, Barman SN, Stone MB, Hughes JB, Shanks JV (2001) Studies on plant-mediated fate of the explosives RDX and HMX. Chemosphere 44: 1259-1264

Boominathan R, Doran PM (2002) Ni-induced oxidative stress in roots of the Ni hyperaccumulator, *Alyssum bertolonii*. New Phytol 156: 205-215

Boominathan R, Doran PM (2003a) Cadmium tolerance and antioxidative defenses in hairy roots of the cadmium hyperaccumulator, *Thlaspi caerulescens*. Biotechnol Bioeng 83: 158-167

Boominathan R, Doran PM (2003b) Organic acid complexation, heavy metal distribution and the effect of ATPase inhibition in hairy roots of hyperaccumulator plant species. J Biotechnol 101: 131-146

Boominathan R, Saha-Chaudhury NM, Sahajwalla V, Doran PM (2004) Production of nickel bio-ore from hyperaccumulator plant biomass: applications in phytomining. Biotechnol Bioeng 86: 243-250

Coniglio MS, Busto VD, González PS, Medina MI, Milrad S, Agostini E (2008) Application of *Brassica napus* hairy root cultures for phenol removal from aqueous solutions. Chemosphere 72: 1035-1042

Cuello JL and Yue LC (2008) Ebb-and-Flow Bioreactor Regime and Electrical Elicitation: Novel Strategies for Hairy Root Biochemical Production. Electronic J Integrative Biosciences, this issue

de Araujo BS, Charlwood VB, Pletsch M (2002) Tolerance and metabolism of phenol and chloroderivatives by hairy root cultures of *Daucus carota* L. Environ Pollut 117: 329-335

de Araujo BS, de Oliveira JO, Machado SS, Pletsch M (2004) Comparative studies of peroxidases from hairy roots of *Daucus carota, Ipomea batatas* and *Solanum aviculare*. Plant Sci 167: 1151-1157

de Araujo BS, Dec J, Bollag JM, Pletsch M (2006) Uptake and transformation of phenol and chlorophenols by hairy root cultures of *Daucus carota*, *Ipomoea batatas* and *Solanum aviculare*. Chemosphere 63: 642-651

Doty SL, James CA, Moore AL, Vajzovic A, Singleton GL, Ma C, Khan Z, Xin G, Kang JW, Park JY, Meilan R, Strauss SH, Wilkerson J, Farin F, Strand SE (2007) Enhanced phytoremediation of volatile environmental pollutants with transgenic trees. Proc Natl Acad Sci USA 104: 16816-16821

Doty SL (2008) Enhancing phytoremediation through the use of transgenics and endophytes. New Phytol 179: 318-333.

Eapen S, Suseelan KN, Tivarekar S, Kotwal SA, Mitra R (2003) Potential for rhizofiltration of uranium using hairy root cultures of *Brassica juncea* and *Chenopodium amaranticolor*. Environ Res 91: 127-133

Gavrilkescu M, Pavel LV, and Cretescu I (2008) Characterization and remediation of soils contaminated with uranium. J Hazard Mater, doi:10.1016/j.hazmats.2008.07.103

Georgiev MI, Pavlov AI, Bley T (2007) Hairy root type plant in vitro systems as sources of bioactive substances. Appl Microbiol Biotechnol 74: 1175-1185 González PS, Capozucca CE, Tigier HA, Milrad SR, Agostini E (2006) Phytoremediation of phenol from wastewater, by peroxidases of tomato hairy root cultures. Enzyme Microbial Technol 39: 647-653

Guillon S, Trémeouillax-Guiller J, Pati PK, Rideau M, Gantet P (2006) Hairy root research: Recent scenario and exciting prospects. Curr Opin Plant Biol 9: 341-346

Gujarathi NP, Haney BJ, Park HJ, Wickramasinghe SR, Linden JC (2005) Hairy roots of *Helianthus annuus*: a model system to study phytoremediation of tetracycline and oxytetracycline. Biotechnol Prog 21: 775-780

Gujarathi NP, Linden JC (2005) Oxytetracycline inactivation by putative reactive oxygen species released to nutrient medium of *Helianthus annuus* hairy root cultures. Biotechnol Bioeng 92: 393-402

Hajeb P, Selemat J, Ismail A, Abu Bakar F, Bakar J, Lioe HN (2008) Hair mercury level of coastal communities in Malaysia: A linkage with fish consumption. Eur Food Res Technol 227: 1349-1355 Harris RC, Rudd JWM, Amyot M, Babiarz CL, Beaty KG, Blanchfield PJ, Bodaly RA, Branfireun BA, Gilmour CC, Graydon JA, Heyes A, Hintelmann H, Hurley JP, Kelly CA, Krabbenhoft DP, Lindberg SE, Mason RP, Paterson MJ, Podemski CL, Robinson A, Sandilands KA, Southworth GR, St. Louis VL, Tate MT (2007) Whole-ecosystem study shows rapid fishmercury response to changes in mercury deposition. Proc Natl Acad Sci USA 104: 16586-16591

Hatcher JM, Delea KC, Richardson JR, Pennell KD, Miller GW (2008) Disruption of dopamine transport by DDT and its metabolites. Neurotoxicol 29: 682-690

Kučerova P, Macková M, Chromá L, Burkhard J, Tříska J, Demnerová K, Macek T (2000) Metabolism of polychlorinated biphenyls by *Solanum nigrum* hairy root clone SNC-90 and analysis of transformation products. Plant Soil 225: 109-115

Kuiper I, Lagendijk EL, Bloemberg GV, Lugtenberg BJ (2004) Rhizoremediation: a beneficial plant-microbe interaction. Mol Plant Microbe Interact 17: 6-15

Macková M, Macek T, Kučerová P, Burkhard J, Pazlarová J, Demnerová K (1997a) Degradation of polychlorinated biphenyls by hairy root culture of *Solanum nigrum*. Biotechnol Lett 19: 787-790

Macková M, Macek T, Ocenaskova J, Burkhard J, Demnerová K, Pazlarová J (1997b) Biodegradation of polychlorinated biphenyls by plant cells. Int Biodeterior Biodegrad 39: 317-325

McAfee BJ, White EE, Pelcher LE, Lapp MS (1993) Root Induction in pine (*Pinus*) and larch (*Larix*) sp. using *Agrobacterium rhizogenes*. Plant Cell Tissue Organ Cult 34: 53-62.

Nedelkoska TV, Doran PM (2000a) Characteristics of heavy metal uptake by plant species with potential for phytoremediation and phytomining. Minerals Eng 13: 549-561

Nedelkoska TV, Doran PM (2000b) Hyperaccumulation of cadmium by hairy roots of *Thlaspi caerulescens*. Biotechnol Bioeng 67: 607-615

Nedelkoska TV, Doran PM (2001) Hyperaccumulation of nickel by hairy roots of *Alyssum* species: Comparison with whole regenerated plants. Biotechnol Prog 17: 752-759

Pilon-Smits E (2005) Phyotoremediation. Annu Rev Plant Biol 56: 15-39

Rentz JA, Alvarez PJJ, Schnoor JL (2005) Benzo [a]pyrene co-metabolism in the presence of plant root extracts and exudates: Implications for phytoremediation. Environ Pollut 136: 477-484

Rezek J, Macek T, Macková M, Tříska J (2007) Plant metabolites of polychlorinated biphenyls in hairy root culture of black nightshade *Solanum nigrum* SNC-90. Chemosphere 69: 1221-1227

Sadasivaiah S, Tozan Y, Breman JG (2007) Dichlorodiphenyltrichloroethylyne (DDT) for indoor residual spraying in Africa: How can it be used for malaria control?. Am J Trop Med Hyg 77: 249-263

Singh S, Melo JS, Eapen S, D' Souza SF (2006) Phenol removal using *Brassica juncea* hairy roots: Role of inherent peroxidase and H<sub>2</sub>O<sub>2</sub>. J Biotechnol 123: 43-49

Srivastava S, Srivastava AK (2007) Hairy root culture for mass-production of high-value secondary metabolites. Crit Rev Biotechnol 27: 29-43

Suresh B, Ravishankar GA (2004) Phytoremediationa novel and promising approach for environmental clean-up. Crit Rev Biotechnol 24: 97-124

Suresh B, Sherkhane PD, Kale S, Eapen S, Ravishankar GA (2005) Uptake and degradation of DDT by hairy root cultures of *Cichorium intybus* and *Brassica juncea*. Chemosphere 61: 1288-1292

Tamaoki M, Freeman JL, Pilon-Smits EAH (2008) Cooperative ethylene and jasmonic acid signaling regulates selenite resistance in Arabidopsis. Plant Physiol 146: 1219-1230

Van Aken B (2008) Transgenic plants for phytoremediation: helping nature to clean up environmental pollution. Trends Biotechnol 26: 225-227

Veena V, Taylor CG (2007) *Agrobacterium rhizogenes*: Recent developments and promising applications. In Vitro Cell Dev Biol 43: 383-403

Villarreal ML, Caspeta L, and Quintero-Ramirez R (2008) From the Mayan highlands to the bioreactors: *In vitro* tissue culture of the Mexican Medicinal plant *Solanum chrysotrichum*. Electronic J Integrative Biosciences, this issue

Vinterhalter B, Savić J, Platiša J, Raspor M, Ninković S, Mitić N, Vinterhalter D (2008) Nickel tolerance and hyperaccumulation in shoot cultures regenerated from hairy root cultures of *Alyssum murale* Waldst et Kit. Plant Cell Tis Organ Cult 94: 299-303

### Electronic Journal of Integrative Biosciences 3(1): 57-65

Special Issue on Hairy Roots (A. Lorence and F. Medina-Bolivar, co-editors) © by Arkansas State University

Walker TS, Bais HP, Halligan KM (2003) Metabolic profiling of root exudates of *Arabidopsis thaliana*. J Agric Food Chem 51: 2548-2554

Weathers P, Liu C, Towler M, and Wyslouzil B (2008) Mist reactors: Principle, comparison of various systems, and case studies. Electronic J Integrative Biosciences, this issue

Wevar-Oller AL, Agostini E, Talano MA, Capozucca C, Milrad SR, Tigier HA, Medina MI (2005)

Overexpression of a basic peroxidase in transgenic tomato (*Lycopersicon esculentum* Mill. cv. Pera) hairy roots increases phytoremediation of phenol. Plant Sci 169: 1102-1111

Xingmao M, Burken JG (2003) TCE Diffusion to the atmosphere in phytoremediation applications. Environ Sci Technol 37: 2534-2539